Articular Cartilage Characterization Using Tantalum and Iodine-based Dual-Contrast Photon-Counting Computed Tomography

Petri Paakkari^{1,2}, Satu I. Inkinen^{3,4}, Jiri Jäntti^{1,2}, Juuso Tuppurainen^{1,2}, Maria C. Fugazzola⁵, Anisha Joenathan⁶, Sampo Ylisiurua^{4,7}, Miika T. Nieminen⁴, Mark W. Grinstaff⁶, Heikki Kröger², Juha Töyräs^{1,2,8}, Juuso T.J. Honkanen², Janne T.A. Mäkelä^{1,2}

¹University of Eastern Finland, Kuopio, Finland, ²Kuopio University Hospital, Kuopio, Finland, ³Helsinki University Hospital, Helsinki, Finland,

⁴University of Oulu, Oulu, Finland, ⁵Utrecht University, Utrecht, The Netherlands, ⁶Boston University, Boston, MA, United States,

⁷Oulu University Hospital, Oulu, Finland, ⁸The University of Queensland, Brisbane, Australia

petri.paakkari@uef.fi

Disclosures: None

INTRODUCTION: Photon-counting detector computed tomography (PCD-CT) is a novel spectral CT imaging technique. PCDs detect individual photons and classify them into fixed energy bins, thus enabling spectral imaging, and e.g., separation of multiple contrast agents from a single scan. In this study, using non-ionic iodinated iodixanol and cationic tantalum oxide nanoparticles (TaO-NPs), we employ a dual-contrast PCD-CT technique to assess structural and biomechanical characteristics of equine articular cartilage.

METHODS: <u>Samples</u>: Intercondylar notch (n = 15) and medial femoral condyle (n = 15) articular cartilage samples (cylindrical plugs, d = 8.5 mm, n = 30) were extracted from equine stifle joints (acquired from an abattoir, N = 15). The plugs were split into quarters: three quarters of each plug were used for reference methods and one quarter of each plug was immersed for 96 h (T = 37 °C) in a dual contrast agent mixture of cationic TaO-NPs (Ta₂O₅, 20 mg·Ta₂O₅/mL) and non-ionic iodixanol (VisipaqueTM, 40 mg·I/mL) (Fig. 1A). Non-articulating surfaces were sealed with cyanoacrylate before the immersion. PCD-CT: The setup consisted of PCD (XC-Flite FX15, XCounter AB), motorized rotator (NR360S, Thorlabs Inc.), and mini-focus X-ray source (IXS1203MF, VJX) with 3 mm aluminum and 0.5 mm copper filters. The used tube voltage was 120 kVp and the used low and total energy bins were 10-80 keV and 10-120 keV, respectively. The samples were imaged before the immersion (0-hour) and at a 96-hour timepoint (Fig. 1B). Calibration of the concentration estimation was based on separately scanned solutions with ten different contrast agent concentrations. Voxel size was $68 \times 68 \times 68$ um³ in the reconstructed volume. Biomechanics: Before splitting the plugs, a four-step stress-relaxation protocol (step size: 4% of remaining cartilage thickness, relaxation: 10 min) and sinusoidal dynamic test (strain amplitude: 4% of remaining cartilage thickness, frequency: 1 Hz, cycles: 4) were conducted with a custom-made biomechanical tester fitted with a 0.25 kg load-cell (Model 31, Honeywell International Inc.) and a flat-ended cylindrical indenter (d = 0.55 mm). Hayes corrected equilibrium modulus, strain-dependent instantaneous modulus, and dynamic modulus were calculated (E_{Eo}, E_{Inst}, and E_{Dyn}, respectively). Proteoglycan content: Spatial proteoglycan (PG) content was determined from Safranin-O-stained sections using digital densitometry. Analysis of CT images: Calibration coefficients were calculated from fits of the measured attenuation values of the calibration solutions. Then a custom-made MATLAB (R2020b, MathWorks) code was used to estimate the contrast agent concentrations in the cartilage of the low and total energy bin reconstructions (Fig. 1B) [1]. Subsequently, depthwise contrast agent partition (i.e., ratio against the original bath concentration) profiles and bulk (full-thickness) cartilage partition values were calculated (Fig. 1C). Statistics: Spearman's correlation coefficients (r) between bulk contrast agent partitions and reference data were calculated with MATLAB.

RESULTS: Concentration estimation was validated with nine different concentration mixtures of contrast agents: mean errors were 18.7% and 8.3% for TaO-NP and iodixanol, respectively. Depthwise partition profiles were determined for all samples and mean (\pm standard deviation) partition profiles were calculated (Fig. 1C). The bulk TaO-NP partition correlated positively with E_{Eq} and E_{Dyn} (Table 1). The bulk iodixanol partition had a strong negative correlation with all the biomechanical moduli. No significant correlation was detected between PGs and the contrast agents alone, but when TaO-NP partition was normalized (divided) with iodixanol partition, a significant positive correlation was detected. Normalization also improved the correlation with biomechanical moduli, and a moderate correlation was observed between normalized TaO-NPs and E_{Inst} .

DISCUSSION: Herein we describe the first use of dual-contrast PCD-CT imaging using TaO-NPs and iodixanol to correlate contrast agent partition with cartilage biomechanics and composition. The dual-contrast method reveals that the partition of cationic TaO-NPs correlates positively with E_{Eq} . This was expected, as cationic contrast agents are known to bind to anionic PGs [1-3], which mainly determine cartilage equilibrium stiffness. Even though the collagen network inhibits TaO-NPs diffusion [2], a positive correlation was found between TaO-NP partition and E_{Dyn} , which can be explained by healthy cartilage and cross-correlation. Iodixanol partition showed a significant negative correlation with all the biomechanical moduli. As it is a non-ionic contrast agent and its diffusion is mainly driven by osmotic pressure, the water content in the cartilage dominates the uptake. The normalization of the TaO-NP partition with the iodixanol diffusion is decreased in the presence of TaO-NPs. Thus, the normalization with iodixanol could be used to improve the sensitivity of TaO-NPs, i.e., iodixanol's faster diffusion might enhance the performance of normalized TaO-NPs in earlier timepoints.

SIGNIFICANCE/CLINICAL RELEVANCE: We demonstrate that the dual-contrast PCD-CT method, utilizing cationic TaO-NPs and non-ionic iodixanol, allows biomechanical and structural evaluation of articular cartilage. Especially, the normalization of the TaO-NPs with iodixanol improves the accuracy of the evaluation.

REFERENCES: [1] Paakkari et al. (2021) Sci. Rep. 11:5556, [2] Jäntti et al. (2022) Osteoarthr. Cartil. 30:S277, [3] Lawson et al. (2021) ACS Nano. 15:19175-19184

ACKNOWLEDGEMENTS: Grants from the Competitive State Research Funding of the Kuopio University Hospital Catchment Area (5063579), Orion Research Foundation, and Maud Kuistila Memorial Foundation are acknowledged.



 Table 1: Spearman's correlation coefficients between bulk

 cartilage
 contrast agent partitions, and biomechanical

 moduli, and proteoglycan (PG) content.

	Iodixanol	TaO-NP	TaO-NP normalized
Equilibrium Modulus (E _{Eq})	-0.488**	0.531**	0.668***
Instantaneous Modulus (E _{Inst})	-0.504**	0.360	0.511**
Dynamic Modulus, 1 Hz (E _{Dyn})	-0.622***	0.555**	0.702***
PG-content	-0.426	0.395	0.552*

Statistical significance: * p<0.05; ** p<0.01; *** p<0.001

Figure 1: A) Cylindrical plugs (n = 30) were extracted from equine stifle joint. Each plug was split to quarters after biomechanical (BM) measurements: one quarter was used for PCD-CT imaging and the rest were used for reference methods, e.g., digital densitometry (DD). B) Low and total energy bin (LE and TE, respectively) reconstructions were used to determine the contrast agent partitions at 96-hour timepoint. C) Depthwise partition profiles for the contrast agents were extracted (I = iodixanol, Ta = tantalum oxide nanoparticles). 0% is the articular cartilage surface and 100% is the cartilage-bone interface.