

QUESTION 3: Is the biofilm on orthopedic implant surface permeable to neutrophils and macrophages in vivo? Are these innate immune cells (meaning any macrophages or neutrophils) capable of engulfing and killing bacteria?

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Response:

A mature bacterial biofilm has limited permeability to neutrophils and macrophages. Those that get through are clinically ineffective at eradicating biofilm bacteria. While neutrophils and macrophages are capable of engulfing and killing planktonic bacteria, they are not innately capable of effectively engulfing and killing sessile bacteria in biofilm.

Level of Evidence: Strong

Delegate Vote: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

Post Meeting Rationale:

The authors completed a systematic review on the peer-reviewed literature identified by a PubMed search performed on February 24, 2018 using the keywords “electron microscopy” and “biofilm” and implant surface. The search identified 148 references from 1991 to 2018. Of these, three references discussed “neutrophil”¹⁻³ and eight references discussed “macrophage”^{1; 3-9}. Another PubMed search with the words “biofilm”, “neutrophils” and “phagocytosis” gave 66 references, and a search with the words “biofilm”, “macrophages” and “phagocytosis” gave 60 references. All these references were reviewed in order to select those that meet the question criteria. Furthermore, to assess the literature for negative findings, a PubMed search was performed using the keywords “natural history” and “biofilm” and “implant,” which identified three references¹⁰⁻¹². Within this literature, only four publications described EM of explanted infected biofilms.

The ability of these immune cells to penetrate mature bacterial biofilms and phagocytosis the infecting bacteria have mainly been evaluated in cystic fibrosis¹⁻⁵, urinary catheter related infection,⁶⁻¹⁴ and periprosthetic infection¹⁵.

Neutrophils (PMN) have shown the ability of sticking, but not penetrating, into a mature biofilm and phagocytizing biofilm encased microorganisms^{1-5,7,8,11,16-20}. The exopolymeric substances of the biofilm matrix seem to be involved in the formation of neutrophil extracellular traps (NETs) in biofilm of *Streptococcus suis*¹⁶, *Candida albicans*⁷ and *C. glabrata*⁸. Data shows that neutrophils can destroy a 2-6 day old *S. aureus* biofilm, but a mature biofilm is capable of resisting penetration by these cells²¹.

Alhede M et al, evaluated the role of immune system against biofilm formed by *Pseudomonas aeruginosa*. They demonstrated that both *in vitro* and *in vivo* biofilms of *P. aeruginosa* produce a shield of excreted rhamnolipids, which offers protection from the bactericidal activity of PMNs²².

Another study showed that PMNs surround biofilm and become activated, but were not able to migrate into the biofilm, probably because of a lack of a chemotactic signal as well as by hindrance of migration into the “slimy” material. Thus, the inability of PMNs to penetrate biofilm results in progression of implant related infections ²³.

Macrophages can penetrate into a mature biofilm in a similar way as neutrophils, and phagocytize biofilm encased microorganisms, but not destroy them ^{6,9,10,15}. Moreover, these sessile phagocytized bacteria can even persist into peri-implant tissue inside macrophage-like cells not only in experimental models, but also in the tissues of patients with intravenous catheters colonized by different bacteria ^{13,14}. *In vivo* model studies in *S. aureus* prosthetic infection showed that limited bacterial macrophage uptake is due to inflammatory attenuation by *S. aureus* biofilm ¹⁰, which favor the transformation from M1 type macrophages presents a high antimicrobial activity to M2 type macrophages with limited antimicrobial activity ¹⁰, and the cell death induction through LukAB ²⁴ and Hla production ¹⁵. At the site of staphylococcus biofilm infection, macrophages exhibit: down-regulation of IL-1 β , TNF α , CXCL2, and CCL2 expression, reduced bacterial uptake, minimal iNOS expression. Consequently, these inhibited macrophages display low efficiency in killing phagocytized bacteria, and reduced induction of lymphocyte produced interferon- γ . As a result, these scavenging cells appear able to migrate into the biofilm, but cannot clear the site from the pathogen causing the infection as their bactericidal activity appears compromised ²³.

References:

1. Hänsch GM, Brenner-Weiss G, Prior B, et al. 2008. The extracellular polymer substance of *Pseudomonas aeruginosa*: too slippery for neutrophils to migrate on? *Int. J. Artif. Organs* 31(9):796–803.
2. Hänsch GM, Prior B, Brenner-Weiss G, et al. 2014. The *Pseudomonas* quinolone signal (PQS) stimulates chemotaxis of polymorphonuclear neutrophils. *J. Appl. Biomater. Funct. Mater.* 12(1):21–26.
3. Jesaitis AJ, Franklin MJ, Berglund D, et al. 2003. Compromised host defense on *Pseudomonas aeruginosa* biofilms: characterization of neutrophil and biofilm interactions. *J. Immunol. (Baltimore, Md. 1950)* 171(8):4329–4339.
4. Parks QM, Young RL, Poch KR, et al. 2009. Neutrophil enhancement of *Pseudomonas aeruginosa* biofilm development: human F-actin and DNA as targets for therapy. *J. Med. Microbiol.* 58(Pt 4):492–502.
5. Takeoka K, Ichimiya T, Yamasaki T, Nasu M. 1998. The in vitro effect of macrolides on the interaction of human polymorphonuclear leukocytes with *Pseudomonas aeruginosa* in biofilm. *Chemotherapy* 44(3):190–197.
6. Hanke ML, Heim CE, Angle A, et al. 2013. Targeting macrophage activation for the prevention and treatment of *Staphylococcus aureus* biofilm infections. *J. Immunol. (Baltimore, Md. 1950)* 190(5):2159–2168.
7. Johnson CJ, Cabezas-Olcoz J, Kernien JF, et al. 2016. The Extracellular Matrix of *Candida albicans* Biofilms Impairs Formation of Neutrophil Extracellular Traps. *PLoS Pathog.* 12(9):e1005884.
8. Johnson CJ, Kernien JF, Hoyer AR, Nett JE. 2017. Mechanisms involved in the triggering of neutrophil extracellular traps (NETs) by *Candida glabrata* during planktonic and biofilm growth. *Sci. Rep.* 7(1):13065.
9. Spiliopoulou AI, Krevvata MI, Kolonitsiou F, et al. 2012. An extracellular *Staphylococcus epidermidis* polysaccharide: relation to Polysaccharide Intercellular Adhesin and its implication in phagocytosis. *BMC Microbiol.* 12:76.
10. Thurlow LR, Hanke ML, Fritz T, et al. 2011. *Staphylococcus aureus* biofilms prevent macrophage phagocytosis and attenuate inflammation in vivo. *J. Immunol. (Baltimore, Md. 1950)* 186(11):6585–6596.
11. Boelens JJ, Dankert J, Murk JL, et al. 2000. Biomaterial-associated persistence of *Staphylococcus epidermidis* in pericatheter macrophages. *J. Infect. Dis.* 181(4):1337–1349.
12. Broekhuizen CAN, de Boer L, Schipper K, et al. 2007. Peri-implant tissue is an important niche for *Staphylococcus epidermidis* in experimental biomaterial-associated infection in mice. *Infect. Immun.* 75(3):1129–1136.

13. Broekhuizen CAN, Schultz MJ, van der Wal AC, et al. 2008. Tissue around catheters is a niche for bacteria associated with medical device infection. *Crit. Care Med.* 36(8):2395–2402.
14. Broekhuizen C a. N, Sta M, Vandenbroucke-Grauls CMJE, Zaat S a. J. 2010. Microscopic detection of viable *Staphylococcus epidermidis* in peri-implant tissue in experimental biomaterial-associated infection, identified by bromodeoxyuridine incorporation. *Infect. Immun.* 78(3):954–962.
15. Scherr TD, Hanke ML, Huang O, et al. 2015. *Staphylococcus aureus* Biofilms Induce Macrophage Dysfunction Through Leukocidin AB and Alpha-Toxin. *MBio* 6(4).
16. Ma F, Yi L, Yu N, et al. 2017. *Streptococcus suis* Serotype 2 Biofilms Inhibit the Formation of Neutrophil Extracellular Traps. *Front. Cell. Infect. Microbiol.* 7:86.
17. Maurer S, Fouchard P, Meyle E, et al. 2015. Activation of Neutrophils by the Extracellular Polymeric Substance of *S. Epidermidis* Biofilms is Mediated by The Bacterial Heat Shock Protein Groel. *J. Biotechnol. Biomater.* 5(1):176–83.
18. Zimmerli W, Lew PD, Cohen HJ, Waldvogel FA. 1984. Comparative superoxide-generating system of granulocytes from blood and peritoneal exudates. *Infect. Immun.* 46(3):625–630.
19. Guenther F, Stroh P, Wagner C, et al. 2009. Phagocytosis of staphylococci biofilms by polymorphonuclear neutrophils: *S. aureus* and *S. epidermidis* differ with regard to their susceptibility towards the host defense. *Int. J. Artif. Organs* 32(9):565–573.
20. Leid JG, Shirtliff ME, Costerton JW, Stoodley P. 2002. Human leukocytes adhere to, penetrate, and respond to *Staphylococcus aureus* biofilms. *Infect. Immun.* 70(11):6339–6345.
21. Hirschfeld J. 2014. Dynamic interactions of neutrophils and biofilms. *J. Oral Microbiol.* 6:26102.
22. Alhede M, Bjarnsholt T, Givskov M, Alhede M. 2014. *Pseudomonas aeruginosa* biofilms: mechanisms of immune evasion. *Adv. Appl. Microbiol.* 86:1–40.
23. Arciola CR, Campoccia D, Speziale P, et al. 2012. Biofilm formation in *Staphylococcus* implant infections. A review of molecular mechanisms and implications for biofilm-resistant materials. *Biomaterials* 33(26):5967–5982.
24. Melehani JH, James DBA, DuMont AL, et al. 2015. *Staphylococcus aureus* Leukocidin A/B (LukAB) Kills Human Monocytes via Host NLRP3 and ASC when Extracellular, but Not Intracellular. *PLoS Pathog.* 11(6):e1004970.