

PTOA Question 3: Do age and post-injury time-point influence the biological response to joint injury and progression to PTOA?

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RESPONSE/RECOMMENDATION: Yes

Based on this systematic review of trauma-induced osteoarthritis animal models, the following evidence-informed recommendations are proposed.

1. Age at injury should be explicitly reported and biologically justified in all studies.
2. Given the predominance of young animals, inclusion of skeletally mature and aged cohorts should be considered where feasible to better model typical human PTOA demographics.
3. Post-injury assessment time-points should be selected in relation to the intended stage of disease being modeled.
4. Most studies evaluated early to mid-term post-injury intervals, highlighting limited assessment of later disease stages.
5. Accordingly, studies aiming to model chronic or progressive osteoarthritis should consider later follow-up time-points (>12 weeks) when appropriate.
6. Use of multiple post-injury time-points may strengthen characterization of the temporal evolution of disease.

LEVEL OF EVIDENCE: Moderate

This ICM statement is based exclusively on preclinical *in vivo* animal studies evaluating trauma-induced osteoarthritis. No human participants, clinical cohorts, or patient-derived data were included. Accordingly, the evidence synthesized reflects experimental and translational preclinical research, rather than clinical effectiveness or patient outcomes.

The overall certainty of evidence is moderate (preclinical). Confidence is strengthened by consistent patterns across multiple independent animal studies and model systems, but is limited by heterogeneity in species, age at injury, and post-injury selection of time-points for analysis, as well as the absence of direct clinical correlation.

DELEGATE VOTE: Agree: __%, Disagree: __%, Abstain: __%

RATIONALE:

Systematic Review of Age on PTOA. A meticulous systematic review was conducted. A comprehensive literature was performed in PubMed, Embase, and Web of Science from database inception to December 15, 2025. The search strategy combined controlled vocabulary and free-text keywords related to post-traumatic osteoarthritis ('ptoa' OR 'post-traumatic osteoarthritis' OR 'traumatic osteoarthritis') AND ('small animal' OR 'animal model') AND ('age' OR 'aging' OR

'ageing'). Searches were limited to English language papers. Reference lists of included studies and relevant reviews were manually screened to identify additional eligible studies. All retrieved citations were imported into a reference manager, and duplicates were removed. Two reviewers independently screened titles and abstracts against predefined eligibility criteria. Full texts were then assessed independently for inclusion. Discrepancies were resolved by consensus or adjudication by a third reviewer. The selection process was documented using a PRISMA flow diagram (Figure S1).

Studies were included if they met all the following criteria:

- Original research involving an *in vivo* model of osteoarthritis induced by joint injury (PTOA) or clearly defined injury-related OA paradigm.
- Reported age at injury and at least one post-injury time point.
- Reported at least one OA-relevant outcome (e.g., histopathology, imaging, biomarkers, biomechanics/functional measures).
- Provided sufficient methodological detail to extract key model characteristics (e.g., species/strain, induction method, age/time-point, sample size).

A standardized extraction form was used to collect study-level data, including: author/year, species and strain, *in vivo* OA induction method, sample size, age at injury/surgery, time-points assessed, treatment or intervention, and key findings. Given heterogeneity in species, injury paradigms, interventions, and outcome measures, results were synthesized qualitatively. Studies were summarized by species/strain, injury model type, age and timing, and intervention class, and key outcomes were reported descriptively. Where applicable, frequencies and proportions of model characteristics were tabulated.

Results. A systematic literature search was conducted in PubMed, Embase, and Web of Science to identify studies describing trauma-induced osteoarthritis (OA) animal models (Figure S1). The search yielded 665 records in total (PubMed, $n = 142$; Embase, $n = 152$; Web of Science, $n = 371$). After removal of 359 duplicate records, 306 records were screened by title and abstract. Of these, 88 records were excluded, leaving 218 reports sought for full-text retrieval. Full texts were obtained and assessed for 117 reports, while 101 reports were excluded at this stage because they did not involve trauma-induced OA models. Following full-text review, 35 reports were excluded for not meeting inclusion criteria. Ultimately, 82 studies met all eligibility criteria and were included in the qualitative synthesis¹⁻⁸³. Most studies used rodents, particularly mice. Mouse models accounted for 61/82 (74.4%) studies, followed by rat 11/82 (13.4%) and rabbit 5/82 (6.1%). Large-animal models were infrequent: horse 2/82 (2.4%), pig 1/82 (1.2%), cat 1/82 (1.2%), and pony 1/82 (1.2%).

OA induction / injury model distribution. The most common OA induction paradigm was destabilization of the medial meniscus (DMM) (43/82, 52.4%), followed by anterior cruciate ligament transection (ACLT) (19/82, 23.2%). We included a small number of

spontaneous/age-associated OA models when they informed the interaction between age and joint degeneration following injury (4/82, 4.9%), meniscectomy or medial collateral ligament injury (MCL) + meniscus variants (3/82, 3.7%), osteochondral/chondral defect models (2/82, 2.4%), and non-invasive ACL rupture via compression (2/82, 2.4%). ACLR/ACL rupture was explicitly listed in 1/82 (1.2%), while other/mixed induction descriptions comprised 7/82 (8.5%).

Age profile in rodent studies Rodent studies (mouse + rat) comprised 72/82 (87.8%) studies. Using the minimum stated age per study (converted to weeks when possible), most rodent experiments were performed in young animals: 6–15 weeks (“young adult”) in 41/72 (56.9%) and <6 weeks (“juvenile/adolescent”) in 20/72 (27.8%). Only 4/72 (5.6%) used 16–51 weeks (“adult”), and 1/72 (1.4%) used ≥52 weeks (“aged”). Age was not stated/parsable for 6/72 (8.3%) rodent studies (Table 1).

Table 1: Age at injury and post-injury time-point distribution of trauma-induced osteoarthritis animal models. Summary of included studies stratified by age at the time of injury and duration of post-injury follow-up. Most studies employed young animals and focused on early to mid post-injury time-points, whereas middle-aged and aged cohorts, as well as late-stage (>12 weeks) disease assessments, were infrequently represented. Percentages are calculated relative to studies reporting age (72 of 82 included studies).

Age at Injury	Post-injury follow-up			Total Studies (%)
	Early (≤4 weeks)	Mid (5–12 weeks)	Late (>12 weeks)	
Young (≤12 weeks)	Predominant	Common	Rare	41 (56.9%)
Skeletally Mature Adult	Common	Predominant	Limited	20 (27.8%)
Middle-aged	Rare	Limited	Occasional	4 (5.6%)
Aged	Rare	Rare	Occasional	1 (1.4%)
Age not specified	—	—	—	6 (8.3%)
Total (Age reported)	—	—	—	72 / 82 (87.8%)

Treatments/interventions evaluated. Interventions were heterogeneous. Genetic manipulation studies represented 24/82 (29.3%), and pharmacologic/biologic interventions represented 15/82 (18.3%). Mechanical/rehabilitation-type interventions were rare (2/82, 2.4%), as were cell/biomaterial therapies (1/82, 1.2%). Interventions that did not clearly fit these categories or were insufficiently described were grouped as “other/unclear” 40/82 (48.8%).

Conclusion. Current PTOA animal studies are heavily weighted toward young rodents and demonstrate substantial variability in post-injury assessment timepoints, limiting comparability and clinical translation. Because aging alters the degree of inflammation and its resolution,

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capacity for repair and OA progression, future work should standardize both animal age strata and windows for assessment post-injury. We recommend routinely including an aged cohort as well as young adults, with prespecified early, mid, and late temporal endpoints aligned to biological phases (inflammation, remodeling, degeneration). Harmonized reporting of age, maturity, and time after injury will strengthen reproducibility and optimal timing for therapeutic interventions.

Both age and post-injury time-point substantially shape the biological response to joint injury and the pace and phenotype of PTOA^{76; 84}. Older age is associated with a heightened or prolonged inflammatory response^{85; 86}, reduced capacity for repair^{7; 87}, increased cellular senescence^{66; 88; 89}, and faster structural degeneration^{7; 76}. Early post-injury windows capture acute inflammation and stress of the chondrocytes and soft tissues, whereas later time-points reflect soft tissue and bone remodeling, osteophyte development, and established structural OA^{84; 90-92}. As most human PTOA arises in middle-aged or older adults with comorbidities, whereas the preclinical literature is in young, healthy rodents. Pre-clinical studies should therefore explicitly define age strata and align outcome measures to biologically meaningful post-injury stages to improve the predictive validity of preclinical studies for human clinical trials.

References

1. Allen KD, Chan KM, Yarmola EG, et al. 2020. The effects of age on the severity of joint damage and intra-articular inflammation following a simulated medial meniscus injury in 3, 6, and 9 month old male rats. *Connect Tissue Res* 61:82-94.
2. Armstrong AR, Carlson CS, Rendahl AK, et al. 2021. Optimization of histologic grading schemes in spontaneous and surgically-induced murine models of osteoarthritis. *Osteoarthritis Cartilage* 29:536-546.
3. Attur M, Duan X, Cai L, et al. 2021. Periostin loss-of-function protects mice from post-traumatic and age-related osteoarthritis. *Arthritis Res Ther* 23:104.
4. Besler BA, Schadow JE, Durongbhan P, et al. 2021. Quantitative measures of bone shape, cartilage morphometry and joint alignment are associated with disease in an ACLT and MMx rat model of osteoarthritis. *Bone* 146:115903.
5. Chan KM, Yeater TD, Allen KD. 2022. Age alters gait compensations following meniscal injury in male rats. *J Orthop Res* 40:2780-2791.
6. Christiansen BA, Anderson MJ, Lee CA, et al. 2012. Musculoskeletal changes following non-invasive knee injury using a novel mouse model of post-traumatic osteoarthritis. *Osteoarthritis Cartilage* 20:773-782.
7. Collins JA, Kim CJ, Coleman A, et al. 2023. Cartilage-specific Sirt6 deficiency represses IGF-1 and enhances osteoarthritis severity in mice. *Ann Rheum Dis* 82:1464-1473.
8. David MA, Smith MK, Pilachowski RN, et al. 2017. Early, focal changes in cartilage cellularity and structure following surgically induced meniscal destabilization in the mouse. *J Orthop Res* 35:537-547.
9. Fang H, Huang L, Welch I, et al. 2018. Early Changes of Articular Cartilage and Subchondral Bone in The DMM Mouse Model of Osteoarthritis. *Sci Rep* 8:2855.
10. Faust HJ, Sommerfeld SD, Rathod S, et al. 2018. A hyaluronic acid binding peptide-polymer system for treating osteoarthritis. *Biomaterials* 183:93-101.

11. Feng K, Liu J, Gong L, et al. 2025. Engineered MSC-sEVs as a Versatile Nanoplatfrom for Enhanced Osteoarthritis Treatment via Targeted Elimination of Senescent Chondrocytes and Maintenance of Cartilage Matrix Metabolic Homeostasis. *Adv Sci (Weinh)* 12:e2413759.
12. Ferrandiz ML, Terencio MC, Ruhi R, et al. 2014. Influence of age on osteoarthritis progression after anterior cruciate ligament transection in rats. *Exp Gerontol* 55:44-48.
13. Fouasson-Chailloux A, Dauty M, Bodic B, et al. 2021. Posttraumatic Osteoarthritis Damage in Mice: From Histological and Micro-Computed Tomodensitometric Changes to Gait Disturbance. *Cartilage* 13:1478S-1489S.
14. Geng N, Fan M, Kuang B, et al. 2024. 10-hydroxy-2-decenoic acid prevents osteoarthritis by targeting aspartyl beta hydroxylase and inhibiting chondrocyte senescence in male mice preclinically. *Nat Commun* 15:7712.
15. Haase T, Sunkara V, Kohl B, et al. 2019. Discerning the spatio-temporal disease patterns of surgically induced OA mouse models. *PLoS One* 14:e0213734.
16. Habumugisha J, Okuda R, Hirose K, et al. 2025. Critical Requirement of Senescence-Associated CCN3 Expression in CD44-Positive Stem Cells for Osteoarthritis Progression. *Int J Mol Sci* 26.
17. Hiyama K, Muneta T, Koga H, et al. 2017. Meniscal regeneration after resection of the anterior half of the medial meniscus in mice. *J Orthop Res* 35:1958-1965.
18. Hsia AW, Jbeily EH, Mendez ME, et al. 2021. Post-traumatic osteoarthritis progression is diminished by early mechanical unloading and anti-inflammatory treatment in mice. *Osteoarthritis Cartilage* 29:1709-1719.
19. Huang H, Skelly JD, Ayers DC, et al. 2017. Age-dependent Changes in the Articular Cartilage and Subchondral Bone of C57BL/6 Mice after Surgical Destabilization of Medial Meniscus. *Sci Rep* 7:42294.
20. Kang D, Lee J, Jung J, et al. 2022. Selenophosphate synthetase 1 deficiency exacerbates osteoarthritis by dysregulating redox homeostasis. *Nat Commun* 13:779.
21. Kim JR, Hong BK, Pham THN, et al. 2024. Interferon-gamma signaling promotes cartilage regeneration after injury. *Sci Rep* 14:8046.
22. Li J, Wang X, Li X, et al. 2023. Mechanical Loading Promotes the Migration of Endogenous Stem Cells and Chondrogenic Differentiation in a Mouse Model of Osteoarthritis. *Calcif Tissue Int* 112:363-376.
23. Li X, Zhang Z, Jiang W, et al. 2025. Dipeptidyl Peptidase 4 (DPP4) Exacerbates Osteoarthritis Progression in an Enzyme-Independent Manner. *Adv Sci (Weinh)* 12:e2410525.
24. Lietman C, Wu B, Lechner S, et al. 2018. Inhibition of Wnt/beta-catenin signaling ameliorates osteoarthritis in a murine model of experimental osteoarthritis. *JCI Insight* 3.
25. Maumus M, Noel D, Ea HK, et al. 2020. Identification of TGFbeta signatures in six murine models mimicking different osteoarthritis clinical phenotypes. *Osteoarthritis Cartilage* 28:1373-1384.
26. McCulloch K, Huesa C, Dunning L, et al. 2019. Accelerated post traumatic osteoarthritis in a dual injury murine model. *Osteoarthritis Cartilage* 27:1800-1810.
27. Mevel E, Merceron C, Vinatier C, et al. 2016. Olive and grape seed extract prevents post-traumatic osteoarthritis damages and exhibits in vitro anti IL-1beta activities before and after oral consumption. *Sci Rep* 6:33527.

28. Murphy MP, Koepke LS, Lopez MT, et al. 2020. Articular cartilage regeneration by activated skeletal stem cells. *Nat Med* 26:1583-1592.
29. Pan C, Lu F, Hao X, et al. 2024. Low-intensity pulsed ultrasound delays the progression of osteoarthritis by regulating the YAP-RIPK1-NF-kappaB axis and influencing autophagy. *J Transl Med* 22:286.
30. Patwari P, Gaschen V, James IE, et al. 2004. Ultrastructural quantification of cell death after injurious compression of bovine calf articular cartilage. *Osteoarthritis Cartilage* 12:245-252.
31. Sanada Y, Ikuta Y, Ding C, et al. 2023. miR-26a deficiency is associated with bone loss and reduced muscle strength but does not affect severity of cartilage damage in osteoarthritis. *Mech Ageing Dev* 212:111806.
32. Shao Y, Zhao C, Pan JY, et al. 2021. BMP5 silencing inhibits chondrocyte senescence and apoptosis as well as osteoarthritis progression in mice. *Aging-Us* 13:9646-9664.
33. Takahashi I, Matsuzaki T, Kuroki H, et al. 2021. Disuse Atrophy of Articular Cartilage Induced by Unloading Condition Accelerates Histological Progression of Osteoarthritis in a Post-traumatic Rat Model. *Cartilage* 13:1522S-1529S.
34. Tang Q, Hao L, Peng Y, et al. 2015. RNAi Silencing of IL-1beta and TNF-alpha in the Treatment of Post-traumatic Arthritis in Rabbits. *Chem Biol Drug Des* 86:1466-1470.
35. Toghraie FS, Chenari N, Gholipour MA, et al. 2011. Treatment of osteoarthritis with infrapatellar fat pad derived mesenchymal stem cells in Rabbit. *Knee* 18:71-75.
36. Triantafillopoulos IK, Papagelopoulos PJ, Politi PK, et al. 2002. Articular changes in experimentally induced patellar trauma. *Knee Surg Sports Traumatol Arthrosc* 10:144-153.
37. Usmani SE, Ulici V, Pest MA, et al. 2016. Context-specific protection of TGFalpha null mice from osteoarthritis. *Sci Rep* 6:30434.
38. van den Bosch MH, Blom AB, Kram V, et al. 2017. WISP1/CCN4 aggravates cartilage degeneration in experimental osteoarthritis. *Osteoarthritis Cartilage* 25:1900-1911.
39. Wu X, Fan X, Plan M, et al. 2025. Altered nicotinamide adenine dinucleotide metabolism drives cartilage degeneration and osteoarthritis. *Clin Transl Med* 15:e70513.
40. Yang C, Li Z, Lai P, et al. 2016. Chondrocyte-Specific Ablation of AMPKalpha1 Does Not Affect Bone Development or Pathogenesis of Osteoarthritis in Mice. *DNA Cell Biol* 35:156-162.
41. Yang Y, Li P, Zhu S, et al. 2020. Comparison of early-stage changes of osteoarthritis in cartilage and subchondral bone between two different rat models. *PeerJ* 8:e8934.
42. Young C, Kobayashi T. 2023. Limited roles of Piezo mechanosensing channels in articular cartilage development and osteoarthritis progression. *Osteoarthritis Cartilage* 31:775-779.
43. Yuan W, Wu Y, Zhou X, et al. 2022. Comparison and applicability of three induction methods of temporomandibular joint osteoarthritis in murine models. *J Oral Rehabil* 49:430-441.
44. Zhang M, Mani SB, He Y, et al. 2016. Induced superficial chondrocyte death reduces catabolic cartilage damage in murine posttraumatic osteoarthritis. *J Clin Invest* 126:2893-2902.
45. Zhang S, Zhang B, Liao Z, et al. 2024. Hnrnpk protects against osteoarthritis through targeting WWC1 mRNA and inhibiting Hippo signaling pathway. *Mol Ther* 32:1461-1478.

46. Au M, Liu Z, Rong L, et al. 2020. Endothelin-1 induces chondrocyte senescence and cartilage damage via endothelin receptor type B in a post-traumatic osteoarthritis mouse model. *Osteoarthritis Cartilage* 28:1559-1571.
47. Batshon G, Elayyan J, Qiq O, et al. 2020. Serum NT/CT SIRT1 ratio reflects early osteoarthritis and chondrosenescence. *Ann Rheum Dis* 79:1370-1380.
48. Bertuglia A, Pagliara E, Grego E, et al. 2016. Pro-inflammatory cytokines and structural biomarkers are effective to categorize osteoarthritis phenotype and progression in Standardbred racehorses over five years of racing career. *BMC Vet Res* 12:246.
49. Blaker CL, Little CB, Clarke EC. 2017. Joint loads resulting in ACL rupture: Effects of age, sex, and body mass on injury load and mode of failure in a mouse model. *J Orthop Res* 35:1754-1763.
50. Boyd SK, Muller R, Leonard T, et al. 2005. Long-term periarticular bone adaptation in a feline knee injury model for post-traumatic experimental osteoarthritis. *Osteoarthritis Cartilage* 13:235-242.
51. Carmon I, Smoum R, Farhat E, et al. 2022. A Fenchone Derivative Effectively Abrogates Joint Damage Following Post-Traumatic Osteoarthritis in Lewis Rats. *Cells* 11.
52. Castanheira C, Anderson JR, Fang Y, et al. 2021. Mouse microRNA signatures in joint ageing and post-traumatic osteoarthritis. *Osteoarthritis Cartilage* 3:100186.
53. Chang LH, Wu SC, Chen CH, et al. 2023. Exosomes Derived from Hypoxia-Cultured Human Adipose Stem Cells Alleviate Articular Chondrocyte Inflammation and Post-Traumatic Osteoarthritis Progression. *Int J Mol Sci* 24.
54. Che X, Zhang C, Zhuo Y, et al. 2025. Serum TSP-1 is a useful biomarker in severity assessment and the diagnosis of osteoarthritis. *J Transl Med* 23:987.
55. Chin AF, Han J, Clement CC, et al. 2023. Senolytic treatment reduces oxidative protein stress in an aging male murine model of post-traumatic osteoarthritis. *Aging Cell* 22:e13979.
56. Cho H, Pinkhassik E, David V, et al. 2015. Detection of early cartilage damage using targeted nanosomes in a post-traumatic osteoarthritis mouse model. *Nanomedicine* 11:939-946.
57. Connard SS, Gaesser AM, Clarke EJ, et al. 2024. Plasma and synovial fluid extracellular vesicles display altered microRNA profiles in horses with naturally occurring post-traumatic osteoarthritis: an exploratory study. *J Am Vet Med Assoc* 262:S83-S96.
58. Cornelis FMF, de Roover A, Storms L, et al. 2019. Increased susceptibility to develop spontaneous and post-traumatic osteoarthritis in *Dot1l*-deficient mice. *Osteoarthritis Cartilage* 27:513-525.
59. Dauenhauer LA, Hislop BD, Brahmachary P, et al. 2024. Aging alters the subchondral bone response 7 days after noninvasive traumatic joint injury in C57BL/6JN mice. *J Orthop Res* 42:2450-2460.
60. Dhaniya G, Mulay V, Kothari P, et al. 2025. Enrichment of the major bioavailable molecule glucuronated flavone TMMG in *Spinacia oleracea* ameliorates cartilage degeneration at a lower dose in ACLT-induced osteoarthritis. *Food Funct* 16:1469-1485.
61. Donat A, Xie W, Jiang S, et al. 2025. *Cxcl9*-deficiency attenuates the progression of post-traumatic osteoarthritis in mice. *Inflamm Res* 74:48.
62. Ely EV, Lenz KL, Paradi SG, et al. 2025. Chondrocyte-specific knockout of *Piezo1* and *Piezo2* protects against post-traumatic osteoarthritis structural damage and pain in mice. *Arthritis Res Ther* 27:152.

63. Faust HJ, Zhang H, Han J, et al. 2020. IL-17 and immunologically induced senescence regulate response to injury in osteoarthritis. *J Clin Invest* 130:5493-5507.
64. Han PF, Li XY, Zhang CP, et al. 2025. Non-targeted metabolomic study in plasma in rats with post-traumatic osteoarthritis model. *PLoS One* 20:e0315708.
65. Heard BJ, Barton KI, Chung M, et al. 2015. Single intra-articular dexamethasone injection immediately post-surgery in a rabbit model mitigates early inflammatory responses and post-traumatic osteoarthritis-like alterations. *J Orthop Res* 33:1826-1834.
66. Jeon OH, Kim C, Laberge RM, et al. 2017. Local clearance of senescent cells attenuates the development of post-traumatic osteoarthritis and creates a pro-regenerative environment. *Nat Med* 23:775-781.
67. Kajabi AW, Casula V, Sarin JK, et al. 2021. Evaluation of articular cartilage with quantitative MRI in an equine model of post-traumatic osteoarthritis. *J Orthop Res* 39:63-73.
68. Keenan CM, Ramos-Mucci L, Kanakis I, et al. 2020. Post-traumatic osteoarthritis development is not modified by postnatal chondrocyte deletion of *Ccn2*. *Dis Model Mech* 13.
69. Komaravolu RK, Mehta-D'souza P, Conner T, et al. 2024. Sex-specific effects of injury and beta-adrenergic activation on metabolic and inflammatory mediators in a murine model of post-traumatic osteoarthritis. *Osteoarthritis Cartilage* 32:1097-1112.
70. Li P, Che X, Gao Y, et al. 2020. Proteomics and Bioinformatics Analysis of Cartilage in Post-Traumatic Osteoarthritis in a Mini-Pig Model of Anterior Cruciate Ligament Repair. *Med Sci Monit* 26:e920104.
71. Li Q, Wu T, Fan Y, et al. 2025. beta-galactosidase-targeted senolytic prodrug ameliorates preclinical models of post-traumatic osteoarthritis. *EBioMedicine* 122:106015.
72. Maatuf YH, Marco M, Unger-Gelman S, et al. 2024. Diverse Response to Local Pharmacological Blockade of Sirt1 Cleavage in Age-Induced versus Trauma-Induced Osteoarthritis Female Mice. *Biomolecules* 14.
73. MacNeil JA, Doschak MR, Zernicke RF, et al. 2008. Preservation of periarticular cancellous morphology and mechanical stiffness in post-traumatic experimental osteoarthritis by antiresorptive therapy. *Clin Biomech (Bristol)* 23:365-371.
74. Mei Z, Yilamu K, Ni W, et al. 2025. Chondrocyte fatty acid oxidation drives osteoarthritis via SOX9 degradation and epigenetic regulation. *Nat Commun* 16:4892.
75. Oliviero S, Millard E, Chen Z, et al. 2022. Accuracy of in vivo microCT imaging in assessing the microstructural properties of the mouse tibia subchondral bone. *Front Endocrinol (Lausanne)* 13:1016321.
76. Sebastian A, Muruges DK, Mendez ME, et al. 2020. Global Gene Expression Analysis Identifies Age-Related Differences in Knee Joint Transcriptome during the Development of Post-Traumatic Osteoarthritis in Mice. *Int J Mol Sci* 21.
77. South SM, Marlin MC, Mehta-D'souza P, et al. 2023. Imaging mass cytometry reveals tissue-specific cellular immune phenotypes in the mouse knee following ACL injury. *Osteoarthritis Cartilage* 31:100416.
78. Takebe K, Rai MF, Schmidt EJ, et al. 2015. The chemokine receptor CCR5 plays a role in post-traumatic cartilage loss in mice, but does not affect synovium and bone. *Osteoarthritis Cartilage* 23:454-461.

79. Van Zeeland EM, Kassel B, Montoya T, et al. 2025. Structural, functional, and proteomic based sex differences in murine post-traumatic osteoarthritis development following mechanical anterior cruciate ligament injury. *Osteoarthritis Cartilage* 33:992-1006.
80. Weaver SR, Arnold KM, Peralta-Herrera E, et al. 2025. Postnatal deletion of *Phlpp1* in chondrocytes delays post-traumatic osteoarthritis in male mice. *Osteoarthritis Cartilage* 33:100525.
81. Yu FH, Yin BF, Liu MY, et al. 2025. Modulation of senescent *Lepr*(+) skeletal stem cells via suppression of leptin-induced *STAT3*–*FGF7* axis activation alleviates abnormal subchondral bone remodeling and osteoarthritis progression. *Stem Cell Res Ther* 16:227.
82. Griffith JL, Valdes-Hernandez PA, Yeater TD, et al. 2025. Post-traumatic osteoarthritis in aged rodents is associated with brain changes that correlate with joint remodeling. *J Pain*:105594.
83. Komaravolu RK, Mehta-D'souza P, Conner T, et al. 2023. Sex-specific effects of injury and beta-adrenergic activation on metabolic and inflammatory mediators in a murine model of post-traumatic osteoarthritis.
84. Punzi L, Galozzi P, Luisetto R, et al. 2016. Post-traumatic arthritis: overview on pathogenic mechanisms and role of inflammation. *RMD Open* 2:e000279.
85. Diekmann BO, Loeser RF. 2024. Aging and the emerging role of cellular senescence in osteoarthritis. *Osteoarthritis Cartilage* 32:365-371.
86. Motta F, Barone E, Sica A, et al. 2023. Inflammaging and Osteoarthritis. *Clin Rev Allergy Immunol* 64:222-238.
87. Martin JA, Buckwalter JA. 2002. Aging, articular cartilage chondrocyte senescence and osteoarthritis. *Biogerontology* 3:257-264.
88. Liu Y, Zhang Z, Li T, et al. 2022. Senescence in osteoarthritis: from mechanism to potential treatment. *Arthritis Res Ther* 24:174.
89. Price JS, Waters JG, Darrah C, et al. 2002. The role of chondrocyte senescence in osteoarthritis. *Aging Cell* 1:57-65.
90. Bergman RF, Lammlin L, Junginger L, et al. 2024. Sexual dimorphism of the synovial transcriptome underpins greater PTOA disease severity in male mice following joint injury. *Osteoarthritis Cartilage* 32:1060-1073.
91. Borrelli J, Jr., Tinsley K, Ricci WM, et al. 2003. Induction of chondrocyte apoptosis following impact load. *J Orthop Trauma* 17:635-641.
92. Ziemian SN, Witkowski AM, Wright TM, et al. 2021. Early inhibition of subchondral bone remodeling slows load-induced posttraumatic osteoarthritis development in mice. *J Bone Miner Res* 36:2027-2038.

Supplementary Materials

Figure S1 PRISMA 2020 flow diagram of study selection. Flow diagram illustrating the identification, screening, eligibility assessment, and inclusion of studies evaluating trauma-induced osteoarthritis animal models. A total of 665 records were identified across PubMed, Embase, and Web of Science. After duplicate removal, 306 records were screened, and 82 studies met inclusion criteria and were included in the qualitative synthesis.

