

The Mohawk homeobox gene represents a marker and osteo-inhibitory factor in cranial suture stem cells

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INTRODUCTION: The regeneration of craniofacial bones of the mammalian skeleton necessitates the action of both intrinsic and extrinsic inductive factors from multiple cell types, which function in a hierarchical and temporal fashion to control the differentiation of osteogenic progenitors. Single-cell transcriptomics of developing mouse cranial suture recently identified a suture mesenchymal progenitor population with previously unappreciated tendon- or ligament-associated gene expression profile. Among teno-ligamentous genes is the transcription factor Mohawk (*Mkx*), an IRX-family homeobox protein involved in vertebrate developmental patterning and critical for tenogenesis. *Mkx* regulates cellular processes such as cell adhesion and migration. *Mkx* has also been implicated in homeostatic maintenance of the teno-ligamentous microenvironments, such as the periodontal ligament and Achilles tendon. However the regulatory role of *Mkx* in other tissues has not been explored. In this study, we used transgenic mouse models and single cell transcriptomics to investigate the role of *Mkx*⁺ cranial suture cells.

METHODS: All experiments were conducted under IACUC approval within the Johns Hopkins University. We engineered *Mkx* reporter mice *Mkx*^{CG}; R26R^{tdT} to examine *Mkx*⁺ cell distribution in the cranial vault. Then, single-cell RNA sequencing (scRNA-Seq) was performed on cells from calvarial bones of uninjured, 7 d, and 28 d post-injury in *Mkx* reporter mice. The transcriptional characteristics of calvarial cells and *Mkx*⁺ cells were analyzed during defect healing. Next, a cell ablation strategy was used to assess the requirement of *Mkx*-expressing cells for bone repair by crossbreeding *Mkx*^{tdT} mice with previously validated iDTR mice (*Mkx*^{tdT/iDTR}). *Mkx*⁺ cell ablation was achieved by local diphtheria toxin (DTX) administration. *Mkx* gene knockout (KO) were performed in *Mkx*^{fl/fl} animals by adenovirus-encoding Cre recombinase (Ad-Cre). Calvarial bone healing was assessed following either *Mkx*⁺ cell ablation in *Mkx*^{tdT} and *Mkx*^{tdT/iDTR} mice or local *Mkx* gene deletion in *Mkx*^{fl/fl} animals over a 28 d period, and bone healing was assessed by high resolution micro computed tomography (microCT), as well as histology and immunohistochemistry for Osteocalcin (OCN) and CD31. Unpaired two-tailed Student t-test was used for a two-group comparison.

RESULTS: We demonstrated that *Mkx* reporter identifies an adult cranial suture resident cell population that gives rise to calvarial osteoblasts and osteocytes during homeostatic conditions (Fig.1). scRNA-Seq data reveal that *Mkx*⁺ suture cells display a stem-like phenotype with expression of teno-ligamentous genes. Bone injury with *Mkx*⁺ cell ablation showed significantly impaired bone healing among *Mkx*^{tdT/iDTR} mice. Micro-CT reconstructions and cross-sectional images demonstrated impaired re-ossification (Fig.2A), including bone volume (BV, 40.2% reduction, Fig.2B), fractional BV (BV/tissue volume (TV), 39.9% reduction, Fig.2C), mean diameter of the bone defect area (36.3% increase, Fig.2D), and bone fractional area (BFA, 36.4% decrease, Fig.2E). The reduction in bone healing in *Mkx*^{tdT/iDTR} animals was further confirmed by OCN immunostaining (Fig.2F, G), and associated with reduced angiogenesis at the healing edge, shown as decreased CD31+ blood vessels (54.3% reduction). Remarkably, *Mkx* gene played a critical role as an osteo-inhibitory factor in cranial suture cells, as *Mkx* knockdown or knockout resulted in increased osteogenic differentiation *in vitro* in cranial suture cells (Fig. 3A, B). Furthermore, localized deletion of *Mkx* *in vivo* resulted in robustly increased calvarial defect repair (Fig.3C-G), including BV (93.7% increase), BV/TV (92.7% increase), mean diameter of the bone defect area (25.6% reduction), and BFA (50.5% increase).

DISCUSSION: In summary, we have identified *Mkx*⁺ cells within the suture mesenchyme with a stem/osteoprogenitor phenotype and a teno-ligamentous gene profile that participates in calvarial bone turnover and bone repair. Depletion of this *Mkx*⁺ progenitor cell population hampered the repair of calvarial defects, yet *Mkx* itself also functions as an osteogenic inhibitor. When *Mkx* was deleted locally, strikingly enhanced cranial bone repair was observed.

SIGNIFICANCE/CLINICAL RELEVANCE: Further characterization of *Mkx*⁺ cells may lead to new insights into the regulatory role of *Mkx* in cranial vault patterning and regeneration, as well as new mechanisms to speed skeletal repair or address craniofacial deformities.

